The impact of certain skin diseases on the lives of those affected tends to be underestimated or even dismissed as simply a “cosmetic problem.” Alopecia areata exemplifies such a condition, owing to its substantial disease burden and its often devastating effects on the patient’s quality of life and self-esteem. Although alopecia areata is one of the most common autoimmune diseases, the pathobiology of this chronic, relapsing hair-loss disorder is not fully understood, and the available therapies are disappointing.

This review summarizes the pathogenesis, clinical presentation, and management of alopecia areata and synthesizes relevant background information concerning the biologic and pathobiologic features of the hair follicle. Currently available evidence suggests that alopecia areata can be considered a T-cell–mediated autoimmune disease in which the gradual loss of protection provided by immune privilege of the normal hair follicle plays an important role.

Epidemiology

Alopecia areata is the most frequent cause of inflammation-induced hair loss, affecting an estimated 4.5 million people in the United States. Depending on ethnic background and area of the world, the prevalence of alopecia areata is 0.1 to 0.2%, with a calculated lifetime risk of 2%. Alopecia areata affects both children and adults and hair of all colors. Although the disorder is uncommon in children under 3 years of age, most patients are relatively young: up to 66% are younger than 30 years of age, and only 20% are older than 40 years of age. There is generally no sex predilection, but more men were found to be affected in one study involving a group of subjects who were 21 to 30 years of age. In a study of 226 Chinese patients with alopecia areata who were 16 years of age, the median age at onset was 10 years, and the male:female ratio was 1.4:1; the disorder was more severe in boys and in those with an onset in early childhood.

Alopecia areata is associated with an increased overall risk of other autoimmune disorders (16%). For example, it is accompanied by lupus erythematosus in 0.6% of patients, vitiligo in 4%, and autoimmune thyroid disease in 8 to 28%.

Normal Hair Growth

It is important to understand normal hair growth and the normal immunobiology of the hair follicle in order to appreciate the changes that occur in alopecia areata and thus its clinical presentation and diagnosis. Hair follicles are the only organs in the human body that undergo extensive, lifelong, cyclic transformation. They switch from a period of very rapid growth, pigmentation, and hair-shaft production (anagen, the active-growth phase, with classification ranging from stages I to VI) to a short, apoptosis-driven phase of organ involution (catagen). After catagen, the hair follicle...
enters a period of relative quiescence (telogen) before it reenters anagen (Fig. 1A).20,21 This regenerative cycle is made possible by an abundance of keratinocyte and melanocyte stem cells located for the most part in the so-called bulge area (Fig. 1B).20,21,23,24 Although hair-follicle cycling and regeneration are stem-cell–dependent,24 hair-shaft production and pigmentation are accomplished by the differentiated progeny of these stem cells. These rapidly proliferating keratinocytes and the pigment-producing melanocytes reside in the anagen hair matrix (Fig. 1A).20,21,23,24 The major target of the inflammatory attack in alopecia areata.25-28

**IMMUNOBIOLOGY OF THE HAIR FOLLICLE**

A crucial immunologic feature of the hair follicle is its creation of a milieu of relative immune privilege that normally renders unlikely an autoimmune attack on intrafollicularly expressed autoantigens.29-31 This relative immune privilege is established mainly by suppression of the surface molecules required for presenting autoantigens to CD8+ T lymphocytes (i.e., major histocompatibility complex [MHC] class I antigens [HLA types A, B, and C] in association with MHC class I–stabilizing β2-microglobulin) and by the generation of an overall immunoinhibitory local signaling milieu.29-32 Although the physiological function of immune privilege with respect to hair follicles is not yet evident, we do know that several autoantigens associated with pigment production are highly immunogenic (as seen in vitiligo and halo nevi). Therefore, one plausible theory is that melanogenesis-associated autoantigens generated during active hair-shaft pigmentation — and perhaps other anagen-associated hair-follicle autoantigens — pose a constitutive risk of attracting autoreactive CD8+ T cells already present.29-32 As with other tissues protected by classic immune privilege (e.g., the anterior chamber of the eye, the central nervous system, and the fetal trophoblast),33,34 down-regulation of MHC class I molecules may serve to reduce the risk that follicle-associated autoantigens will be presented to CD8+ T cells.29,30 (See Table 1 in the Supplementary Appendix, available with the full text of this article at NEJM.org.)

This down-regulation of MHC class I molecules, however, entails the risk that the hair follicle may be attacked by natural killer (NK) cells, since NK cells are primed to recognize and eliminate MHC class I–negative cells.35 To reduce this risk, healthy hair follicles appear to down-regulate the expression of ligands that stimulate the activation of NK-cell receptors (NKG2D)36 and secrete molecules that inhibit NK-cell and T-cell functions, such as transforming growth factors β1 and β2, α melanocyte–stimulating hormone, and macrophage migration inhibitory factor.32-37

In the hair follicles of healthy rodents, immune privilege generated in this manner is so effective that even transplanted allogeneic melanocytes escape rejection if they manage to migrate into the hair bulb during anagen (Table 1 in the Supplementary Appendix).30,38

**CLINICAL PRESENTATION AND DIAGNOSIS**

Alopecia areata is manifested as the loss of hair in well-circumscribed patches of normal-appearing skin, most commonly on the scalp (Fig. 2 and 3) and in the region of the beard (Fig. 3A).17,39-41 The onset is typically rapid, and the disease can progress to the point where all the hair is lost on the scalp (alopecia areata totalis) or even on the whole body (alopecia areata universalis) (Fig. 2A, 2B, and 2C). Variants of this disorder include ophiasis, in which hair loss affects the occipital
**A**

**Normal Hair Cycle**

- **Epidermis**
- **Connective-tissue sheath**
- **Bulb**
- **Dermal papilla**
- **Sebaceous gland**
- **Hair**

**Anagen**

**Catagen**

**Telogen**

**Return to anagen**

**Active melanogenesis**

---

**B**

**Hair Cycle in Alopecia Areata**

- **Bulge area**
- **Club hair**
- **Old club hair**

**Anagen**

**Catagen**

**Telogen**

**Return to anagen**

- **Melanocyte**
- **Natural killer cell**
- **CD8+ T cell**
- **CD4+ T cell**
- **Mast cell**
- **Dendritic cell**

**Premature induction of catagen**

**Active melanogenesis**
scalp (Fig. 2D); diffuse forms of alopecia (Fig. 2E); and “sudden graying,” a variant in which pigmented hair follicles are attacked, with the result that preexisting gray hairs are demasked (Fig. 2F). These presentations, together with tell-tale clinical signs, such as exclamation-mark hairs (Fig. 3B), cadaver hairs (Fig. 3C), nail pitting (Fig. 3D), and the growth of white hair in formerly alopecic lesions (Fig. 3E), often render the diagnosis straightforward (Table 1). An association of patchy hair loss with autoimmune disorders, as well as with atopic dermatitis (in 39% of cases), further points to the correct diagnosis.

If the diagnosis is not clear after a clinical evaluation (Table 1 and Fig. 3), as can be the case with the diffuse variant of alopecia areata, skin biopsy is usually diagnostic. In acute alopecia areata, histologic examination reveals a characteristic “beeswarm pattern” of dense, perifollicular lymphocytic infiltrates around anagen hair follicles (Fig. 1 in the Supplementary Appendix); in patients with chronic disease, this pattern may be absent. (Additional clinical pictures and recommendations for the diagnostic workup are available from the National Alopecia Areata Foundation at www.naaf.org/site/PageServer?pagename=about_alopecia_types.)

MANAGEMENT

Although diagnosing alopecia areata is usually easy, treating it is not. Curative therapy does not exist, and there is a paucity of well-conducted, long-term, controlled trials evaluating therapy for alopecia areata and its effect on the quality of life. Given the often unsatisfactory results of current therapy, some clinicians rely on the high rate of spontaneous remission and will recommend a wig if remission does not occur. Still, limited but often helpful therapeutic options do exist for both acute and chronic, relapsing alopecia areata.

Clinicians have two principal management options: use of an immunosuppressive regimen (preferable for patients with acute and rapidly progressing alopecia areata) or an immune-deviation strategy that manipulates the intracutaneous inflammatory milieu (favored for patients with the chronic, relapsing form). At present, only two approaches reach the level of evidence-based medicine: intrallesional injections of glucocorticoids and the induction of contact allergy.

The best-tested immunosuppressive treatment consists of intradermal injections of triamcinolone acetonide (5 to 10 mg per milliliter) given every 2 to 6 weeks. This agent stimulates localized regrowth in 60 to 67% of cases. Side effects include pain, localized skin atrophy, and depigmentation, and relapses are frequent after treatment has
beaten discontinued. Potent topical glucocorticoids are also widely used, especially in children and in adults with less than 50% loss of scalp hair. High-potency topical glucocorticoids with occlusive dressings are most beneficial and lead to improvement in more than 25% of affected patients; however, glucocorticoid-induced folliculitis is a common adverse effect.

The use of systemic glucocorticoids is limited mainly because of their adverse-event and side-effect profiles. In one study, moderate regrowth of hair (31 to 60%) was observed in 30% of patients treated with oral prednisolone (200 mg given once a week for 3 months); however, in 25% of patients with a response, relapse occurred within 3 months. The best response was achieved with high-dose intravenous methylprednisolone (500 mg given on 3 consecutive days); 147 of 218 patients (67%) treated for multifocal alopecia areata had more than 50% regrowth of hair. However, relapse occurred within a year in one third of those who had a response, and the number of relapses increased with time. Other potential immunosuppressive strategies are suggested by case reports of patients with alopecia areata in conjunction with other autoimmune diseases for whom hair regrowth was complete when systemic immunosuppressant agents such as azathioprine were used to treat the other conditions.

The simplest form of topical immunomodulation therapy is the irritant dithranol (anthralin), an antipsoriatic agent. Dithranol (0.2 to 0.8%) can be applied for 20 to 30 minutes daily as the initial, short-contact therapy, with the length of contact gradually increased by 10 minutes every 2 weeks to a maximum of 1 hour or until a low-grade der-
matitis develops. This treatment deserves consideration as a second-line therapy for adults and children with persistent disease. Indeed, with this approach, hair regrowth was observed in 75% of patients with limited disease (including ophiasis) and in 25% of those with alopecia areata totalis.

The most effective form of immunotherapy is topical sensitization with diphenylcyclopropenone (diphencyprone [not approved by the Food and Drug Administration]) or squaric acid dibutylester. Diphencyprone can now be considered first-line therapy for alopecia areata totalis. First, the patient is sensitized to this obligatory synthetic allergen over a period of 1 to 2 weeks; then the lowest diphencyprone concentration that causes mild irritation is applied weekly (with the concentration subsequently increased to maintain a mild contact dermatitis). Although published results of this treatment vary greatly, one relatively large trial (involving 148 patients) showed hair regrowth in 17% of patients with alopecia areata totalis or universalis, 60% of patients with 75 to 99% hair loss, and all patients with less than 50% hair loss.

Diphencyprone has also been used in children with severe alopecia areata (with regrowth reported in 27 to 33% of cases).

Relapses are common after therapy is discontinued. The most frequent adverse events include pruritus, pain, lymphadenopathy due to local inflammation, generalized contact eczema, influenza-like symptoms, and changes in skin color at the site of allergen application.

Topical minoxidil, a potassium-channel facilitator that has long been used as a general hair-growth stimulant in androgenetic alopecia, can also be used in alopecia areata, ideally in conjunction with other treatments, such as dithranol cream or oral glucocorticoids. After a 6-week course of oral glucocorticoids, topical application of 2% minoxidil may help prevent or delay re-

**Table 1. Diagnostic Criteria for Alopecia Areata.**

<table>
<thead>
<tr>
<th>Diagnostic Tool</th>
<th>Diagnostic Findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Family history</td>
<td>Atopy, thyroid disease, or other autoimmune disorders may be associated with alopecia areata; a family history of any of these disorders may therefore be diagnostic</td>
</tr>
<tr>
<td>Physical examination</td>
<td></td>
</tr>
<tr>
<td>Hair and skin</td>
<td>Most characteristic diagnostic finding is the presence of circumscribed, hairless patches or large alopecic areas in otherwise normal-appearing skin areas; pigmented hair is preferentially attacked and lost in active disease, whereas regrowth is frequently characterized by tufts of white hair; sudden pseudowhitening of hair is observed in a rare, rapidly progressing, diffuse variant form of alopecia areata</td>
</tr>
<tr>
<td>Nails</td>
<td>Nail changes, if present, are usually characterized by pitting; onychodystrophy is less common</td>
</tr>
<tr>
<td>Eyes</td>
<td>Ocular abnormalities include lens opacities and abnormalities of retinal pigment epithelium</td>
</tr>
<tr>
<td>Dermoscopy</td>
<td>Yellow dots (i.e., keratotic plugs in follicular ostia) are often seen in alopecia areata but are not specific for the diagnosis</td>
</tr>
<tr>
<td>Cadaver hairs</td>
<td>Comedo-like cadaver hairs (black dots) may also be present</td>
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<tr>
<td>Exclamation-mark hair</td>
<td>Distal segment of the hair shaft is broader than its proximal end, resembling an exclamation mark</td>
</tr>
<tr>
<td>Follicular ostia</td>
<td>Openings in the hair follicles through which the hair fiber emerges from the skin; these ostia are well preserved in alopecia areata, in contrast to the findings in scarring alopecia</td>
</tr>
<tr>
<td>Pull test*</td>
<td>A positive pull test at the margins of alopecic lesions that produces telogen (“club”) or dystrophic anagen hairs supports a clinical working diagnosis</td>
</tr>
<tr>
<td>Laboratory tests</td>
<td>None of the available tests will confirm the diagnosis, but thyroid-function tests and tests for thyroid antibodies may be advisable because of the increased association between alopecia areata and thyroid autoimmunity; abnormal results of thyroid-function tests, the presence of thyroid autoantibodies, or both further support a clinical or histologic working diagnosis of alopecia areata</td>
</tr>
<tr>
<td>Histologic examination†</td>
<td>Biopsy specimens should be obtained only if the clinical diagnosis is in doubt; on histologic examination, a dense, perifollicular lymphocytic infiltrate is seen in acute alopecia areata</td>
</tr>
</tbody>
</table>

* The pull test is also positive in other conditions, such as fungal hair infections.
† This perifollicular infiltrate can be deceptively subtle in long-standing, chronic disease.
lapse in patients who have had a response to glucocorticoids. Several other topical and systemic agents have been tried, but they have not been shown to offer a clear therapeutic benefit.

**PATHOBIOLOGY OF ALOPECIA AREATA**

Improved pathobiologic concepts may ultimately pave the way to better management and outcomes in alopecia areata. It is important to note that this is a disorder of hair-follicle cycling in a dual sense: inflammatory cells attack only anagen hair follicles, which are then prematurely propelled into the catagen phase (Fig. 1B). Because of inflammation-induced dystrophy of the follicle, the hair shaft can no longer be firmly anchored in the hair canal and is rapidly shed; however, the hair follicle retains its capacity to regenerate and continue cycling, since in alopecia areata — unlike scarring alopecias — hair-follicle stem cells are generally not destroyed. Thus, the loss of hair in this disorder is, in principle, reversible.

Like most other autoimmune diseases, alopecia areata is a chronically relapsing inflammatory disorder, which suggests a cyclic recurrence of disease-promoting events. Also, in the absence of a perifollicular infiltrate, there is no hair loss. The main therapeutic challenge, therefore, is to reduce the already established inflammatory infiltrates and to prevent both recurrence and spread to previously unaffected hair follicles. Unfortunately, currently available therapies do not predictably and satisfactorily meet this challenge.

A better understanding is needed of how the perifollicular infiltrate in alopecia areata develops and why it chiefly forms around follicles in those stages of the hair cycle during which pigment is produced (i.e., anagen III through VI). Alopecia areata may be considered an organ-specific autoimmune disease, since it exclusively affects hair follicles, nails, and (in some patients) the retinal pigment epithelium. Therefore, antigens or autoantigens that are preferentially or exclusively presented in these selected tissues could be important in the pathobiology of this disease. In addition, systemic interferon alfa therapy and tumor necrosis factor α antagonists, which are used to treat other autoimmune diseases, can trigger or aggravate alopecia areata, suggesting that selected cytokines may also be important pathogenetic factors.

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**GENETIC COMPONENT IN ALOPECIA AREATA**

The development of alopecia areata has a strong genetic component (Section I in the Supplementary Appendix). For example, many patients with a family history of alopecia areata also have a personal or family history of atopy, Down’s syndrome, autoimmune polyendocrinopathy–candidiasis–ectodermal dystrophy syndrome, other autoimmune diseases, or a combination of these disorders. Familial cases of alopecia areata, as compared with sporadic cases, are often characterized by a poorer prognosis, more rapid progression, more frequent relapses, and greater resistance to therapy. Relatives of affected family members are also at greatly increased risk for alopecia areata. Substantial ethnic variations in the incidence and relative risk of alopecia areata further underscore the prominent role of genetic factors in its pathogenesis.

In a genomewide association study of 20 families with alopecia areata, Martinez-Mir et al. identified at least four susceptibility loci on chromosomes 6, 10, 16, and 18; a validation set was not included. On chromosome 6, one susceptibility locus was found at 6p, a site that corresponds to the HLA locus; a second locus was found at 6q23.3, a site that is outside the HLA gene cluster. The region on chromosome 16 overlaps with a region near a susceptibility locus for Crohn’s disease. The susceptibility locus for alopecia areata on chromosome 18p also contains a psoriasis-susceptibility region. Indeed, alopecia areata can coexist with psoriasis.

Cytotoxic T-lymphocyte–associated antigen 4 (CTLA4), a costimulatory molecule that is involved in the negative regulation of T-cell activation and has been implicated in psoriasis, may also be a susceptibility gene for alopecia areata — namely, in patients with a severe form of the disorder. The CTLA4 association is supported by another genomewide association study in which Petukhova et al. affirmed the importance of both innate immunity and acquired immunity in the pathogenesis of alopecia areata and underscored the fact that this disorder shares pathways with other autoimmune diseases (Section I in the Supplementary Appendix).

The two genomewide association studies used different approaches to analyze susceptibility loci.
for alopecia areata. Whereas Martinez-Mir et al. scanned the genome within families of patients affected by alopecia areata, Petukhova et al. evaluated 1054 patients and 3278 control subjects and identified 139 single-nucleotide polymorphisms that were significantly associated with alopecia areata. Petukhova et al. first performed an in silico analysis, which was then used to study samples from patients, whereas the study by Martinez-Mir et al. was solely an in silico analysis. Both studies identified susceptibility loci common to alopecia areata on chromosomes 6p (HLA), 6q (UL 16 binding protein [ULBP]), 10p (IL2RA), and 18p (PTPN22). In addition, Petukhova et al. identified some genes that may be associated with alopecia areata and other autoimmune diseases, such as the genes for ULBP, which encode a class of ligands for activating NKG2D (Section I in the Supplementary Appendix).

### Basic Immunopathology

Insights into the immunopathological mechanisms in alopecia areata may be best gained from examining the skin lesions. Although CD4+ T cells predominate numerically in the perifollicular infiltrates, CD8+ T cells appear to be the first lymphocytes to enter the proximal follicular epithelium (Fig. 1B, 1C, and 1E in the Supplementary Appendix).\(^7\) In addition, the numbers of NK cells and mast cells are greatly increased in the perifollicular infiltrates, raising the question of whether these cells are also involved in the pathogenesis of alopecia areata.\(^37,78\) Autoantibodies against follicular autoantigens are often found in the serum and skin of patients with alopecia areata,\(^79,80\) but there is no evidence that they are pathogenic.\(^81\)

In fact, in murine models of alopecia areata, the disease can be transferred by CD8+ T cells alone,\(^7\) especially after the T cells have been primed by contact with melanogenesis-related autoantigens.\(^82,83\) The transfer of CD8+ T cells together with CD4+ T cells is most effective in initiating the disease in the most widely used murine model,\(^7\) whereas the transfer of serum or autoantibodies from patients with alopecia areata fails to elicit hair loss.\(^81\) Conversely, depleting CD8+ T cells restores hair growth in a rat model of alopecia areata.\(^84\) Thus, it is reasonable to consider alopecia areata a CD8+ T-cell–dependent, organ-specific autoimmune disease (Table 2 in the Supplementary Appendix).

### Hypothetical Pathogenesis of Alopecia Areata

It has been hypothesized that alopecia areata develops in a previously healthy hair follicle because its constitutive immune privilege collapses.\(^30,85\) According to this hypothesis, alopecia areata can occur in a genetically predisposed person only when proinflammatory signals (e.g., interferon-γ and substance P) known to up-regulate ectopic MHC class Ia expression in human hair-follicle epithelium\(^32,87\) expose previously “sequestered” follicle–associated autoantigens to preexisting autoreactive CD8+ T cells (Fig. 4). If costimulatory signals and help from other cells, such as CD4+ T cells\(^30,85\) and mast cells,\(^88\) are provided, the lymphocytic infiltrates could attack the hair follicle. Since only anagen hair follicles are attacked, the autoantigens in question may be generated and presented only during anagen (e.g., melanogenesis-related sequence A [MICA] protein,\(^37\) IL2RA,\(^85,89\) and ULBP3 protein).\(^85-87\) This scenario is supported by extensive evidence derived from mouse models of alopecia areata.\(^7-9,82-84,86,90\)

Genomewide association studies suggest that other proinflammatory factors and NK-cell–stimulating ligands may also be active at some stage during the development of alopecia areata\(^37,74\) (Fig. 4, and Sections I and II and Table 2 in the Supplementary Appendix). NK cells and NKG2D and their endogenous ligands have been implicated in the pathogenesis of alopecia areata. Although very few NK cells are observed around healthy anagen hair follicles,\(^79\) lesional follicles show prominent aggregates of CD56+ and NKG2D+ NK cells.\(^37\) Moreover, hair follicles in alopecia areata overexpress MHC class I polypeptide–related sequence A (MICA) protein,\(^37\) a key NKG2D agonist,\(^36\) whereas MICA expression in healthy hair follicles is much more limited.\(^37\) The conclusion that excessive NKG2D-mediated signaling may contribute to the pathogenesis of alopecia areata is underscored by the genetic association between the disease and NKG2D-activating ligands from the MICA family — namely, ULBP3. (ULBP3 protein expression is actually up-regulated around lesional hair follicles in alopecia areata.\(^74\))

### Future Therapy

Current pathobiologic concepts may inform preclinical research to develop better therapeutic options for alopecia areata. Treatment strategies that
Figure 4. Theory of Immune Privilege Collapse in Alopecia Areata.

Under normal conditions, the hair follicle is protected by a relatively labile immune privilege. Patients with a certain specific genetic background are predisposed to abnormalities in the microenvironment of the follicle, allowing follicular autoantigens (e.g., melanogenesis-associated antigens) to be presented to preexisting autoreactive CD8+ T cells. When various costimulatory circumstances occur during anagen (e.g., trauma, infection, or stress), the clinical phenotype of alopecia areata results. (For a more detailed explanation of this process, see the additional description of Figure 4 in the Supplementary Appendix.) CGRP denotes calcitonin gene–related peptide, IDO indoleamine 2,3-dioxygenase, IGF-1 insulin-like growth factor 1, IK down-regulator of HLA II, MHC major histocompatibility complex, MICA MHC class I polypeptide–related sequence A, MIF migration inhibitory factor, αMSH α melanocyte–stimulating hormone, NK natural killer, NKG2D activating receptor for NK cells and subgroups of T lymphocytes, and TGF-β1 transforming growth factor β1.

restores or prevent the collapse of hair-follicle immune privilege and that antagonize excessive NKG2D-mediated signaling or the interaction of pathogenic CD8+ T cells with MHC class I–presented hair-follicle autoantigens may eventually lead to more effective management of this disease.9,19,37,41,91 New therapeutic strategies now being explored in preclinical research are described in Section III in the Supplementary Appendix.

This common autoimmune disorder already provides an excellent, easily accessible model of disease with which to investigate general principles concerning the generation, maintenance, collapse, and restoration of immune privilege.9,30,37,92 In-sights obtained from such research may thus also become relevant to the treatment of other autoimmune diseases characterized by the collapse of immune privilege, such as multiple sclerosis, immune abortion, and autoimmune uveitis.9,33,34,41,92

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